

The role of ELABELA in targeted treatment of acute myeloid leukemia

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Cite this article: Yanardağ Açık D, Karabulut ZT. The role of ELABELA in targeted treatment of acute myeloid leukemia. *J Curr Hematol Oncol Res.* 2025;3(3):67-73.

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Received: 16/05/2025

Accepted: 24/10/2025

Published: 08/08/2025

ABSTRACT

Acute myeloid leukemia is a clonal stem cell disease. Various molecular targets are being sought in this disease which has a low chance of cure. However, it is desired that targeted treatments should have few side effects. We tried to understand whether the embryogenic peptide ELABELA, which has been closely associated with many pathways involved in cancer mechanism, is a peptide worth investigating in targeted treatment of acute myeloid leukaemia.

Keywords: ELABELA, acute myeloid leukemia, cellular pathways, targeted therapy, new peptide

INTRODUCTION AND OVERVIEW OF ACUTE MYELOID LEUKEMIA (AML)

Acute myeloid leukemia (AML) is a genetically, epigenetically and clinically heterogeneous disease characterized by the accumulation and expansion of immature myeloid cells in the bone marrow (BM) and peripheral blood (PB), resulting in failure of normal hematopoiesis.¹

Morphological heterogeneity of AML arises from the diversity of genotypes seen.²

As in other cancers, various intracellular and extracellular signaling pathways, proteins, peptides, receptors etc. in these pathways play a role in the pathogenesis of AML. Although blocking and inhibition of some molecules involved in these complex pathways have led to important developments in cancer treatment, the results are still unsatisfactory.³⁻⁵

CANCER STEM CELLS (CSCS) AND THEIR ROLE IN AML

Cancer stem cells (CSCs) are responsible for cancer recurrence, metastasis and resistance to treatment, especially in leukemias. The activities of CSCs are controlled by many intracellular and extracellular factors and these factors can be used as drug targets for cancer therapy. CSCs have been initially identified in leukemia. CSCs can generate tumors through self-renewal and differentiation into multiple cellular subtypes.⁶⁻¹¹

ELABELA: A NOVEL PEPTIDE IN EMBRYOGENESIS AND CANCER

During embryogenesis, apela (ELABELA/ELA/Toddler) and apelin receptor (AR) mRNA expression is increased, leading to apela-AR signaling that facilitates the migration of progenitor cells to the anterior lateral plate mesoderm via Nodal/TNF β -dependent signaling for proper cardiovascular development in zebrafish.¹²⁻¹⁴ However, following this, apela levels rapidly decrease and apelin mRNA levels increase.¹⁵ CSCs and normal stem cells share some of the same regulatory signaling pathways such as Wnt/ β -catenin, Sonic Hedgehog (Hh), and Notch pathways involved in the self-renewal process.¹⁶⁻¹⁸ Targeting these common pathways may result in complications such as profound cytopenia and BM failure. Therefore, finding targets that have lost their functions after the embryonic period, but reappear or persist in possible CSCs will be advantageous in destroying the cancer stem cell, and even if this does not happen, it will be advantageous in protecting against the side effects of treatment. One of these targets may be Apela peptide, which has lost its function after the embryonic period but is abundantly expressed in embryonic stem cells¹²⁻¹⁴ and has been shown to be expressed in some cancer cells in recent studies.¹⁹⁻²¹

Apela binds to an unknown receptor different from apelin in ESCs, inhibits apoptosis of stem cells and stimulates self-renewal. Apela may have varying effects on ESCs derived from different species. Human and murine pluripotent ESCs respond differently to exogenous apela treatment. In human ESCs, it inhibits apoptosis.²² In mice, apela can cause apoptosis in a non-peptidic manner.²³

In this paper, we examine whether ELABELA, a peptide that has an important role in signaling pathways involved in AML mechanism and especially in ESC, is worth investigating in targeted treatment.

Wnt signaling is required for the sustainability of leukemic stem cells. Several molecules involved in or modulating Wnt signaling have a prognostic value in AML. These include: β -catenin, LEF-1, phosphorylated-GSK3 β , PSMD2, PPAR δ , XPNPEP, sFRP2, RUNX1, AXIN2, PCDH17, CXXC5, LLGL1, and PTK7.²

From a successful treatment perspective, the cancer stem cell model implies that the LSC population must be eliminated to eradicate the disease and achieve long-term remissions.²⁴

Notch-, Wnt- and Hedgehog signaling pathways are particularly important for LSC biology.²⁵

Numerous lines of evidence suggest that abnormal WNT signaling may also impair cancer immune surveillance, thereby stimulating immune escape and resistance to various immunotherapies, including immune checkpoint blockers.^{26,27}

Both p53 and the Wnt signaling pathway play important roles in regulating the differentiation of mouse embryonic stem cells (mESCs). Lee et al.²⁷ identified the Wnt signaling pathway as one of the main targets of p53 in mESCs.

Glycogen synthase kinase-3 (GSK-3) and protein phosphatase 2A (PP2A) are two important signaling molecules involved in AML pathogenesis and MYC stability. GSK-3 β is involved in the regulation of cell metabolism, cell differentiation, apoptosis, autophagy and tumorigenesis of leukemia. PP2A is a serine/threonine phosphatase that suppresses tumor growth and regulates kinase-directed intracellular signaling pathways.²⁹

Takam Kamga et al.²⁹ have shown that pharmacological interference via small molecule inhibitors of Wnt and/or GSK-3 signaling reduces AML cell survival by sensitizing leukemia cells to chemotherapeutic agents both in vitro and in vivo.

The G protein-coupled chemokine receptor CXCR4 is specifically associated with cancer metastasis and HIV-1 infection.³¹ The chemokine receptor, CXCR4, with its ligand chemokine ligand 12 (CXCL12), plays a key role in the survival and migration of normal and malignant stem cells into the BM. High CXCR4 expression in AML and ALL blasts has been shown to be an indicator of poor prognosis for these diseases. Several small molecule inhibitors, short peptides, antibodies and antibody drug conjugates have been developed to more effectively target and kill malignant cells expressing CXCR4. Apelin and apela are two peptide ligands for a class A G-protein coupled receptor (GPCR) called the AR (AR/APJ/APLNR). They function by binding to this receptor, and this is known as the Apelinergic system (Apelin/APJ system). Binding of both endogenous peptides to the AR produces similar physiological effects. The

AR transmembrane (TM) domain is highly homologous to the chemokine receptors CXCR4 and CCR5.^{15,32}

Apelin peptides (apelin and ELABELA) and the GPCR of the same origin, known as the apelin receptor (Aplnr), play key roles in apoptosis, cell proliferation, angiogenesis, metabolic disorders and various cancers.^{33,34} The CXCR4 receptor interacts with CXCL12 (SDF-1) generated by BM endothelial and stromal cells, including CXCL12-bol reticular (CAR) cells. Therefore, CXCR4 may be an attractive molecular target for the treatment of these malignancies. Studies have shown that increased CXCR4 expression is correlated with poor results in patients with AML and B-cell ALL. This is potentially due to the role of CXCR4 in retaining leukemic cells within the BM and their role in activating pathways that support the survival, growth and chemotherapy resistance of these malignant cells. In AML cases, leukemic blasts with high CXCR4 expression have been associated with higher relapse rates and lower overall survival. Patients with FMS-like tyrosine kinase-3 internal tandem duplication (FLT3-ITD) or nucleophosmin mutation have significantly higher CXCR4 expression than their wild-type counterparts. The CXCR4 receptor plays a crucial role in leukemia maintenance through several mechanisms, including survival signaling and retention of leukemia cells in the protective BM niche.³⁵⁻⁴⁴ LSCs are tumor-initiating cells that are resistant to standard chemotherapy and promote the persistence and relapse of AML.⁴⁵

CXCL12 antagonists are designed to disrupt the CXCR4/CXCL12 axis and sensitize malignant cells to chemotherapy.³⁵ Of the CXCL12 antagonists, successful treatment has only been achieved with CX-01.⁴⁶

In apoptosis, the B-cell lymphoma protein 2 (Bcl-2)-associated X (Bax) protein releases cytochrome c from mitochondria, activating a cascade of reactions that assists the successive activation of caspases and ultimately leads to cell death. Bcl-2 is believed to inhibit Bax's release of cytochrome c, thereby restricting the downstream activation of the apoptotic machinery. Both Bax and Bcl-2 are cytoplasmic proteins.⁴⁷ BCL2 is thought to mediate resistance to standard treatment in patients with unfavorable risk AML.⁴⁸

Mutation of the tumor suppressor gene TP53 is associated with poor prognosis in AML. The TP53 gene is largely a stress response protein with functions ranging from apoptosis to cell cycle control. The fate of the p53 protein in the cell is mainly determined by the rate of degradation rather than the rate of generation. p53 is mainly degraded by the interaction of p53 with MDM2 (double minute 2 protein). This coupling of p53/MDM2 can be disrupted when stress or DNA damage is detected, resulting in the stabilization of p53. Overexpression of MDM2 leads to deactivation of p53.^{49,50} MDM2 and MDM4 are negative regulators of TP53 activity targeting degradation via ubiquitination. In AML, MDM2 is frequently overexpressed, so cells can have low functional TP53 activity without TP53 deletion/mutation. Nutlin-3a is a small molecular inhibitor of MDM2/p53 interaction, thus this therapeutic provides increased p53 levels secondary to reduced p53 degradation and consequently up-regulates

apoptosis.^{50,51} It has been suggested that Venetoclax plus idasanutlin may be an effective treatment for relapsed/refractory AML.⁵²

Apelin and apela are two peptide ligands for a class A GPCR called the AR. They function by binding to this receptor, and this is known as the apelinergic system (apelin/APJ system). Coupling of both endogenous peptides to the AR produces similar physiological effects. It is well known that the apelin/APJ system can regulate apoptosis in various cell types, subsequently mediating the occurrence and development of associated diseases. Recent evidence suggests that the Apelin/APJ system affects apoptosis in various diseases through different signaling pathways in different disease pathways. Pretreatment of cardiomyocytes with a pelin-13 effectively prevents apoptosis induced by glucose deprivation (GD) and can significantly increase Akt and mTOR phosphorylation by positive regulation of Bcl-2 protein and negative regulation of Bax and cleaved caspase-3. The apelin/APJ system also increases the expression of Bcl-2 and decreases the expression of Bax protein.⁵³⁻⁵⁷ Inhibition of the apela or apelinergic system will show the effect of both venetoclax and idasanutlin together.

Apela functions as an endogenous ligand for the GPCR, APLNR, and Apela and APLNR have been shown to direct angioblast migration to control vascular patterning in zebrafish embryos.^{14,58,59}

MOLECULAR MECHANISMS LINKING ELABELA TO AML

Apela is predicted to bind to a different receptor than apela.¹⁵ Apela is highly expressed in human blastocysts before implantation and contributes to human embryonic stem cell (hESC) pluripotency through an alternative receptor. In a study in human pluripotent embryonic stem cells (ESCs), apela inhibited cell apoptosis and stimulated self-renewal. However, these undifferentiated stem cells did not express AR. Only after differentiation, about three days later, AR expression was detected.²² This is highly suggestive of the existence of an additional, as yet unknown receptor for apela and indicates that apela is involved in pluripotent stem cells prior to and independent of AR formation. In this case, targeting apela in the treatment of AML, or at least in the preparatory regimen for hematopoietic stem cell transplantation, will perhaps enable cancer stem cell targeting.

The non-coding region of Apela has been shown to be involved in the regulation of p53-mediated DNA damage-induced apoptosis in mouse embryonic stem cells (mESCs). Apela negatively regulates the interaction between heterogeneous nuclear ribonucleoprotein L (hnRNPL) and p53.²³ Seo et al.⁵⁹ showed that DNA damage-induced hnRNP L positively regulates p53 expression. Li et al.²² have shown that Apela negatively regulates the interaction between hnRNPL and p53, resulting in an antiapoptotic effect. Yi et al.¹⁹ showed that apela expression was high in ovarian cancer cells. Disruption of apela expression in these cell lines suppressed cell growth, cell migration and cell cycle progression. They showed that apela demonstrated this effect independently of APLNR and affected cell growth and cell cycle progression

in a p53-dependent way. Loss of apela in high p53-expressing cells caused a decrease in cell number due to cell death, which was caused by p53-induced cell apoptosis.

Mouse double minute2 (MDM2) is a critical negative regulator of the tumour suppressor p53, which plays a key role in controlling its transcriptional activity, protein stability and nuclear localization. MDM2 expression is upregulated in many cancers, resulting in loss of p53-dependent activities such as apoptosis and cell cycle arrest.⁶¹

The PI3K/Akt signaling pathway has been shown to play a critical role in tumorigenesis of hematopoietic cells. Activation of the PI3K/Akt pathway occurs even in the early stages of tumors and is correlated with poor prognosis and therapeutic resistance in various human cancers.^{22,62}

Mammalian target of rapamycin (mTOR) is an evolutionarily conserved kinase in eukaryotes that plays a critical role in sensing and responding to factors such as nutrient availability, energy sufficiency, stress, hormones and mitogens. mTOR forms two complexes, referred to as mTOR complex 1 (mTORC1) and mTORC2. mTORC1 also regulates mitochondrial biogenesis and autophagy.⁶³⁻⁶⁶ Hoshii et al.⁶⁶ in their study revealed that mTORC1 is required for leukemia initiation.

The phosphatidylinositol-3-kinase (PI3K)/Akt/mTOR pathway are two pathways that are crucial for many aspects of cell growth and survival in cancer as well as physiological.⁶⁸

GSK3 β functions as a link between the PI3K/mTOR and WNT/ β -catenin pathways through AMP-activated protein kinase (AMPK).^{69,70}

AMPK activation leads to inhibition of mTORC1 by two independent mechanisms: activation of the negative mTORC1 regulator tuberous sclerosis complex 2 (TSC2) and inhibition of the mTORC1 sub-unit RAPTOR.⁷¹⁻⁷³ c-Myc is an essential transcription factor that plays a convincing role in leukemogenesis. In vitro and in vivo studies have suggested that higher expression of c-Myc is correlated with the progression of AML.⁷⁴ Two key regulators of c-Myc are glycogen synthase kinase 3 β (GSK-3 β) and PP2A.²⁹ GSK-3 β is involved in the regulation of cell metabolism, cell differentiation, apoptosis, and autophagy and leukemia tumorigenesis.⁷⁵ GSK-3 β is also involved in leukemia survival, chemotherapy resistance and body protection.⁷⁶ GSK-3 β is a vital tumor suppressor.⁷⁷ In its tumor suppressor function, GSK-3 β regulates Wnt signaling to passivate inhibitory phosphorylation of β -catenin and prevents cell cycle progression.⁷⁸ Soulet et al.⁷⁸ demonstrated the effects of ELA on autophagy and mTORC1. They showed that ELA is also involved in the regulation of the expression of mTORC1 network proteins and autophagy. They identified ELA as an important positive regulator of mTORC1.

Apela stimulates hESC cell cycle progression and protein translational by activating PI3K/AKT/mTORC1 signaling and blocks stress-induced apoptosis. These pathways are the main signals reported to correlate with apoptosis. MDM2 also inhibits p53 through this pathway. It is predicted that the apelinergic system may inhibit apoptosis through these

common pathways (53- 57). hnRNPc is a negative regulator of p53. It was also shown that the 1-41 p53 region, the region where p53 binds to MDM2, interacts with hnRNPc. These results showed that hnRNPc may be synergistic with MDM2 in regulating p53 stability. Doxorubicin competes with p53 for binding to the RNA recognition motif of hnRNPc, thereby increasing p53 stability and inducing p53-dependent apoptosis.⁸⁰ Chemokines are produced by cancer-associated fibroblasts, a component of stromal cells, and affect the metastatic potential and region-specific spread of cancer cells. Stromal-derived factor-1 (SDF-1/CXCL12) belongs to the CXC chemokine family. The effects of CXCL12 in many cancers have been described, including its role in promoting local invasion and distant lung cancer metastasis.⁸¹⁻⁸³ Wang et al.⁸³ in their study showed that CXCL12 blocks apoptosis in human adenocarcinoma cell line via CXCR4. They observed that the expression levels of Bcl-2 and bcl-xl in the adenocarcinoma cell line were increased by treatment with CXCL12 and decreased by CXCR4 antagonist and JAK2 inhibitor. Apela, which has been shown to be antiapoptotic, has also been shown to interfere with the chemokine CXCR4a signaling pathway.²²

In summary, apela and apelinergic system has been shown to inhibit apoptosis in many steps (via bcl-2, bcl-xl, mdm2, hnRNPc, p53, PI3K/Akt/mTORC1, chemokines). Based on these results, we can say that apela and the apelinergic system occupy a central position in leukaemia.

AKT, ERK and mTOR kinases form a complex pathway that regulates various cellular functions, most prominently cell growth, migration and survival.⁷⁹

AML LSCs (leukemia stem cells) maintain naturally high mitophagy activity through AMP-activated protein kinase (AMPK) activation to maintain a low physiological state of reactive oxygen species (ROS), which is critically required for the preservation of their self-renewal potential.⁸⁵

Autophagy is frequently activated in cancer and is largely associated with metabolic reprogramming and oncogenesis. Autophagy (i.e. lipophagy) has been shown to participate in lipid catabolism to support OXPHOS, i.e. to maintain energy metabolism by delivering free fatty acids (FFAs) to mitochondria via degradation of lipid droplets (LD) in AML cells but not in normal hematopoietic cells. Autophagy supports OXPHOS, whereas inhibition of OXPHOS reduces autophagic flow.⁸⁶

Autophagy is thought to inhibit cancer development. Conversely, once cancer has formed, increased autophagic flow often provides tumor cell survival and growth.⁸⁷⁻⁸⁹ In other words, autophagy has different effects on cancer formation and progression. Therefore, both inhibition and activation of autophagy can be used in treatment.^{87,88}

mTORC1 stimulates biosynthetic pathways, including the synthesis of proteins, lipids and nucleotides, and promotes cell growth by inhibiting cellular catabolism through suppression of the autophagic pathway. However, inhibition of autophagy initiates cancer formation. So mTORC1 probably contributes to the development of AML. Altman et al.⁹⁰ have demonstrated the antileukemic effects of blocking mTORC1

and mTORC2 in cell culture studies. However, mTORC1 negatively regulates and inhibits autophagy. Inhibition of autophagy in AML is among the treatment targets.⁹⁰⁻⁹²

Soulet et al.⁷⁸ showed that ELA plays a role in the regulation of the expression of mTORC1 network proteins. They showed that ELA activates mTORC1 and this activation inhibits autophagy. They also showed that the effect of ELA on autophagy is abolished in the absence of mTORC1 expression. ELA has been shown to be an important positive regulator of mTORC1. This correlation may suggest that mTORC1 and ELA may be jointly responsible for the initiation of AML.

BCL-2 inhibitors (venetoclax) inhibit oxidative phosphorylation (OXPHOS) in leukemic stem cells (LSC) and lead to cell death. Inhibition of OXPHOS reduces autophagic flow.⁸⁶

ELABELA AND POTENTIAL THERAPEUTIC TARGETING IN AML

Targeting apela, which seems to be almost central to the cancer pathways involved in AML, is promising. However, cell culture studies will be very valuable to see the real effect of apela and will reveal more real results. Although blocking/inhibiting Apela seems logical according to the pathways it is involved in and the results of the studies, it is also possible to encounter a different result in cell culture studies. For example, Soulet et al.⁷⁸ reported that sunitinib and ELA showed synergistic effects in suppressing tumor growth and angiogenesis in mice. In the same study, they found that APELA expression was upregulated in colon, lung, stomach and thymoma cancers, but did not change in other cancer tissues analyzed except kidney cancer tissues, while APLN and APLNR expression was upregulated in clear cell renal carcinoma (ccRCC) and downregulated in papillary renal cell carcinoma (RCC). In other words, the expression of apela and APLNR differs in different cancer types; therefore, inhibition or induction of APLNR is likely to show different effects in different cancer types. Sunitinib (SU11248) is a small molecule FLT3 inhibitor with selectivity for PDGFR, VEGFR1, VEGFR2, KIT, and FLT3. It has both direct anti-tumor and antiangiogenic properties. The use of sunitinib is currently approved for the treatment of renal cell carcinoma, gastrointestinal stromal tumor and AML.⁹³ Cell culture studies will be valuable to understand whether this synergistic effect is observed in AML.

CONCLUSION

As a result, many molecules and cancer pathways involved in AML formation have a close connection with Apela. Therefore, Apela seems to be a peptide worth investigating in AML treatment. Cell culture studies and later phase studies will provide valuable information to the literature.

ETHICAL DECLARATIONS

Referee Evaluation Process

Externally peer-reviewed.

Conflict of Interest Statement

The authors have no conflicts of interest to declare.

Financial Disclosure

The authors declared that this study has received no financial support.

Author Contributions

All of the authors declare that they have all participated in the design, execution, and analysis of the paper, and that they have approved the final version.

REFERENCES

- Vago L, Gojo I. Immune escape and immunotherapy of acute myeloid leukemia. *J Clin Invest*. 2020;130(4):1552-1564. doi:10.1172/JCI129204
- Gruszka AM, Valli D, Alcalay M. Wnt signalling in acute myeloid leukaemia. *Cells*. 2019;8(11):1403. doi:10.3390/cells8111403
- Daver N, Schlenk RF, Russell NH, Levis MJ. Targeting FLT3 mutations in AML: review of current knowledge and evidence. *Leukemia*. 2019;33(2):299-312. doi:10.1038/s41375-018-0357-9
- Molica M, Breccia M, Foa R, Jabbour E, Kadia TM. Maintenance therapy in AML: the past, the present and the future. *Am J Hematol*. 2019;94(11):1254-1265. doi:10.1002/ajh.25620
- Kayser S, Levis MJ. Advances in targeted therapy for acute myeloid leukaemia. *Br J Haematol*. 2018;180(4):484-500. doi:10.1111/bjh.15032
- Ajani JA, Song S, Hochster HS, Steinberg IB. Cancer stem cells: the promise and the potential. *Semin Oncol*. 2015;42(Suppl 1): S3-17. doi:10.1053/j.seminoncol.2015.01.001
- Yang L, Shi P, Zhao G, et al. Targeting cancer stem cell pathways for cancer therapy. *Signal Transduct Target Ther*. 2020;5(1):8. doi:10.1038/s41392-020-0110-5
- Visvader JE, Lindeman GJ. Cancer stem cells: current status and evolving complexities. *Cell Stem Cell*. 2012;10:717-728. doi:10.1016/j.stem.2012.05.007
- Lapidot T, Sirard C, Vormoor J, et al. A cell initiating human acute myeloid leukemia after transplantation into SCID mice. *Nature*. 1994;367:645-648. doi:10.1038/367645a0
- Croker AK, Allan AL. Cancer stem cells: implications for the progression and treatment of metastatic disease. *J Cell Mol Med*. 2008;12(2):374-390. doi:10.1111/j.1582-4934.2007.00211.x
- Chng SC, Ho L, Tian J, Reversade B. ELABELA: a hormone essential for heart development signals via the apelin receptor. *Dev Cell*. 2013;27(6): 672-680. doi:10.1016/j.devcel.2013.11.002
- Deshwar AR, Chng SC, Ho L, Reversade B, Scott IC. The apelin receptor enhances Nodal/TGFβ signaling to ensure proper cardiac development. *Elife*. 2016;5:e13758. doi:10.7554/eLife.13758
- Pauli A, Norris ML, Valen E, et al. Toddler: an embryonic signal that promotes cell movement via apelin receptors. *Science*. 2014;343(6172): 1248636. doi:10.1126/science.1248636
- Shin K, Kenward C, Rainey JK. Apelinergic system structure and function. *Compr Physiol*. 2017;8(1):407-450. doi:10.1002/cphy.c170028
- Kanwar SS, Yu Y, Nautiyal J, Patel BB, Majumdar AP. The Wnt/beta-catenin pathway regulates growth and maintenance of colonospheres. *Mol Cancer*. 2010;9:212. doi:10.1186/1476-4598-9-212
- Li C, Heidt DG, Dalerba P, et al. Identification of pancreatic cancer stem cells. *Cancer Res*. 2007;67(3):1030-1037. doi:10.1158/0008-5472.CAN-06-2030
- Wang J, Wakeman TP, Lathia JD, et al. Notch promotes radioresistance of glioma stem cells. *Stem Cells*. 2010;28(1):17-28. doi:10.1002/stem.261
- Acik DY, Bankir M, Baylan FA, Aygun B. Can ELABELA be a novel target in the treatment of chronic lymphocytic leukaemia? *BMC Cancer*. 2019;19(1):1086. doi:10.1186/s12885-019-6325-6
- Yi Y, Tsai SH, Cheng JC, et al. APELA promotes tumour growth and cell migration in ovarian cancer in a p53-dependent manner. *Gynecol Oncol*. 2017;147(3):663-671. doi:10.1016/j.ygyno.2017.10.016
- Ganguly D, Cai C, Sims MM, et al. APELA expression in glioma, and its association with patient survival and tumor grade. *Pharmaceuticals (Basel)*. 2019;12(1):45. doi:10.3390/ph12010045
- Ho L, Tan SY, Wee S, et al. ELABELA is an endogenous growth factor that sustains hESC self-renewal via the PI3K/AKT pathway. *Cell Stem Cell*. 2015;17(4):435-447. doi:10.1016/j.stem.2015.08.010
- Li M, Gou H, Tripathi BK, Huang J. An apela RNA-containing negative feedback loop regulates p53-mediated apoptosis in embryonic stem cells. *Cell Stem Cell*. 2015;16(6):669-683. doi:10.1016/j.stem.2015.04.002
- Dick JE. Acute myeloid leukemia stem cells. *Ann N Y Acad Sci*. 2005;1044:1-5. doi:10.1196/annals.1349.001
- Heidel FH, Arriba-Tutusa P, Armstrong SA, Fischer T. Evolutionarily conserved signaling pathways: acting in the shadows of acute myelogenous leukemia's genetic diversity. *Clin Cancer Res*. 2015;21:240-248. doi:10.1158/1078-0432.CCR-14-1436
- Han Y, Liu D, Li L. PD-1/PD-L1 pathway: current researches in cancer. *Am J Cancer Res*. 2020;10(3):727-742.
- Galluzzi L, Spranger S, Fuchs E, López-Soto A. WNT signaling in cancer immunosurveillance. *Trends Cell Biol*. 2019;29(1):44-65. doi:10.1016/j.tcb.2018.08.005
- Lee KH, Li M, Michalowski AM, et al. A genomewide study identifies the Wnt signaling pathway as a major target of p53 in murine embryonic stem cells. *Proc Natl Acad Sci USA*. 2010;107(1):69-74. doi:10.1073/pnas.0909734107
- Mudgapalli N, Nallasamy P, Chava H, et al. The role of exosomes and MYC in therapy resistance of acute myeloid leukemia: challenges and opportunities. *Mol Aspects Med*. 2019;70:21-32. doi:10.1016/j.mam.2019.10.001
- Takam Kamga P, Dal Collo G, Cassaro A, et al. Small molecule inhibitors of microenvironmental Wnt/β-catenin signaling enhance the chemosensitivity of acute myeloid leukemia. *Cancers (Basel)*. 2020;12(9):2696. doi:10.3390/cancers12092696
- Wu B, Chien EY, Mol CD, et al. Structures of the CXCR4 chemokine GPCR with small-molecule and cyclic peptide antagonists. *Science*. 2010;330(6007):1066-1071. doi:10.1126/science.1194396
- Vermeire K, Schols D, Bell TW. Inhibitors of HIV infection via the cellular CD4 receptor. *Curr Med Chem*. 2006;13(7):731-743. doi:10.2174/092986706776055599
- Chapman NA, Dupré DJ, Rainey JK. The apelin receptor: physiology, pathology, cell signalling, and ligand modulation of a peptide-activated class A GPCR. *Biochem Cell Biol*. 2014;92(6):431-440. doi:10.1139/bcb-2014-0072
- Uribealago I, Hoffmann D, Zhang Y, et al. Apelin inhibition prevents resistance and metastasis associated with anti-angiogenic therapy. *EMBO Mol Med*. 2019;11(8):e9266. doi:10.15252/emmm.201809266
- Cancilla D, Rettig MP, DiPersio JF. Targeting CXCR4 in AML and ALL. *Front Oncol*. 2020;10:1672. doi:10.3389/fonc.2020.01672
- Spoo AC, Lübbert M, Wierda WG, Burger JA. CXCR4 is a prognostic marker in acute myelogenous leukemia. *Blood*. 2007;109:786-791. doi:10.1182/blood-2006-05-024844
- Burger JA, Bürkle A. The CXCR4 chemokine receptor in acute and chronic leukaemia: a marrow homing receptor and potential therapeutic target. *Br J Haematol*. 2007;137:288-296. doi:10.1111/j.1365-2141.2007.06590.x
- Konoplev S, Lin P, Yin CC, et al. CXC chemokine receptor 4 expression, CXC chemokine receptor 4 activation, and wild-type nucleophosmin are independently associated with unfavorable prognosis in patients with acute myeloid leukemia. *Clin Lymphoma Myeloma Leuk*. 2013;13: 686-692. doi:10.1016/j.clml.2013.05.013
- Mannelli F, Cutini I, Gianfaldoni G, et al. CXCR4 expression accounts for clinical phenotype and outcome in acute myeloid leukemia. *Cytometry B Clin Cytom*. 2014;86(5):340-349. doi:10.1002/cytob.21156
- Cao T, Jiang N, Liao H, Shuai X, Su J, Zheng Q. The FLT3-ITD mutation and the expression of its downstream signaling intermediates STAT5 and Pim-1 are positively correlated with CXCR4 expression in patients with acute myeloid leukemia. *Sci Rep*. 2019;9(1):12209. doi:10.1038/s41598-019-48687-z
- Cao T, Ye Y, Liao H, et al. Relationship between CXC chemokine receptor 4 expression and prognostic significance in acute myeloid leukemia. *Medicine*. 2019;98(23):e15948. doi:10.1097/MD.00000000000015948
- Du W, Lu C, Zhu X, et al. Prognostic significance of CXCR4 expression in acute myeloid leukemia. *Cancer Med*. 2019;8(15):6595-6603. doi:10.1002/cam4.2535
- Mohle R, Schittenhelm M, Failenschmid C, et al. Functional response of leukaemic blasts to stromal cell-derived factor-1 correlates with preferential expression of the chemokine receptor CXCR4 in acute myelomonocytic and lymphoblastic leukaemia. *Br J Haematol*. 2000;110(3):563-572. doi:10.1046/j.1365-2141.2000.02157.x
- Abraham M, Pereg Y, Bulvik B, et al. Single dose of the CXCR4 antagonist BL-8040 induces rapid mobilization for the collection of human CD34(+) cells in healthy volunteers. *Clin Cancer Res*. 2017;23(22):6790-6801. doi:10.1158/1078-0432.CCR-16-2919
- Carter JL, Hege K, Kalpage HA, et al. Targeting mitochondrial respiration for the treatment of acute myeloid leukemia. *Biochem Pharmacol*. 2020;182:114253. doi:10.1016/j.bcp.2020.114253

45. Kovacovics TJ, Mims A, Salama ME, et al. Combination of the low anticoagulant heparin CX-01 with chemotherapy for the treatment of acute myeloid leukemia. *Blood Adv.* 2018;2(4):381-389. doi:10.1182/bloodadvances.2017013391
46. Kulsoom B, Shamsi TS, Afsar NA, Memon Z, Ahmed N, Hasnain SN. Bax, Bcl-2, and Bax/Bcl-2 as prognostic markers in acute myeloid leukemia: are we ready for Bcl-2-directed therapy? *Cancer Manag Res.* 2018;10:403-416. doi:10.2147/CMAR.S154608
47. Pelcovits A, Niroula R. Acute myeloid leukemia: a review. *R I Med J* (2013). 2020;103(3):38-40.
48. Bode AM, Dong Z. Post-translational modification of p53 in tumorigenesis. *Nat Rev Cancer.* 2004;4(10):793-805. doi:10.1038/nrc1455
49. George B, Kantarjian H, Baran N, Krockner JD, Rios A. TP53 in Acute myeloid leukemia: molecular aspects and patterns of mutation. *Int J Mol Sci.* 2021;22(19):10782. doi:10.3390/ijms221910782
50. Vassilev LT, Vu BT, Graves B, et al. In vivo activation of the p53 pathway by small-molecule antagonists of MDM2. *Science.* 2004;303(5659):844-848. doi:10.1126/science.1092472
51. Droegemeier K. BCL2/MDM2 inhibitor combo effective in AML. *Cancer Discov.* 2019;9(2):156. doi:10.1158/2159-8290.CD-NB2019-003
52. Liu J, Liu M, Chen L. Novel pathogenesis: regulation of apoptosis by Apelin/APJ system. *Acta Biochim Biophys Sin (Shanghai).* 2017;49(6):471-478. doi:10.1093/abbs/gmx035
53. Zhang Z, Yu B, Tao GZ. Apelin protects against cardiomyocyte apoptosis induced by glucose deprivation. *Chin Med J.* 2009;122(19):2360-2365. doi:10.3760/cma.j.issn.0366-6999.2009.19.031
54. Xin M, Deng X. Nicotine inactivation of the proapoptotic function of Bax through phosphorylation. *J Biol Chem.* 2005;280(11):10781-10789. doi:10.1074/jbc.M500084200
55. Burgering BM, Medema RH. Decisions on life and death: FOXO Forkhead transcription factors are in command when PKB/Akt is off duty. *J Leukoc Biol.* 2003;73(6):689-701. doi:10.1189/jlb.1202629
56. Mayo LD, Donner DB. The PTEN, Mdm2, p53 tumor suppressor network. *Trends Biochem Sci.* 2002;27(9):462-467. doi:10.1016/s0968-0004(02)02166-7
57. Hassan AS, Hou J, Wei W, Hoodless PA. Expression of two novel transcripts in the mouse definitive endoderm. *Gene Expr Patterns.* 2010;10(2-3):127-134. doi:10.1016/j.gexp.2010.02.001
58. Choe W, Albright A, Sulcove J, et al. Functional expression of the seven-transmembrane HIV-1 co-receptor APJ in neural cells. *J Neurovirol.* 2000;6(Suppl 1):S61-69.
59. Seo JY, Kim DY, Kim SH, et al. Heterogeneous nuclear ribonucleoprotein (hnRNP) L promotes DNA damage-induced cell apoptosis by enhancing the translation of p53. *Oncotarget.* 2017;8(31):51108-51122. doi:10.18632/oncotarget.17003
60. Oliner JD, Saiki AY, Caenepeel S. The role of MDM2 amplification and overexpression in tumorigenesis. *Cold Spring Harb Perspect Med.* 2016;6(6):a026336. doi:10.1101/cshperspect.a026336
61. Kawauchi K, Ogasawara T, Yasuyama M, Otsuka K, Yamada O. The PI3K/Akt pathway as a target in the treatment of hematologic malignancies. *Anticancer Agents Med Chem.* 2009;9(5):550-559. doi:10.2174/187152009788451851
62. Ma XM, Blenis J. Molecular mechanisms of mTOR-mediated translational control. *Nat Rev Mol Cell Biol.* 2009;10(5):307-318. doi:10.1038/nrm2672
63. Ramanathan A, Schreiber SL. Direct control of mitochondrial function by mTOR. *Proc Natl Acad Sci USA.* 2009;106(52):22229-22232. doi:10.1073/pnas.0912074106
64. Cunningham JT, Rodgers JT, Arlow DH, Vazquez F, Mootha VK, Puigserver P. mTOR controls mitochondrial oxidative function through a YY1-PGC-1 α transcriptional complex. *Nature.* 2007;450(7170):736-740. doi:10.1038/nature06322
65. Rabinowitz JD, White E. Autophagy and metabolism. *Science.* 2010;330(6009):1344-1348. doi:10.1126/science.1193497
66. Hoshii T, Tadokoro Y, Naka K, et al. mTORC1 is essential for leukemia propagation but not stem cell self-renewal. *J Clin Invest.* 2012;122(6):2114-2129. doi:10.1172/JCI62279
67. Porta C, Paglino C, Mosca A. Targeting PI3K/Akt/mTOR signaling in cancer. *Front Oncol.* 2014;4:64. doi:10.3389/fonc.2014.00064
68. Shakoory A, Ougolkov A, Yu ZW, et al. Deregulated GSK3 β activity in colorectal cancer: its association with tumor cell survival and proliferation. *Biochem Biophys Res Commun.* 2005;334(4):1365-1373. doi:10.1016/j.bbrc.2005.07.041
69. Park YL, Kim HP, Cho YW, et al. Activation of WNT/ β -catenin signaling results in resistance to a dual PI3K/mTOR inhibitor in colorectal cancer cells harboring PIK3CA mutations. *Int J Cancer.* 2019;144(2):389-401. doi:10.1002/ijc.31662
70. Inoki K, Zhu T, Guan KL. TSC2 mediates cellular energy response to control cell growth and survival. *Cell.* 2003;115(5):577-590. doi:10.1016/s0092-8674(03)00929-2
71. Gwinn DM, Shackelford DB, Egan DF, et al. AMPK phosphorylation of raptor mediates a metabolic checkpoint. *Mol Cell.* 2008;30(2):214-226. doi:10.1016/j.molcel.2008.03.003
72. Herzig S, Shaw RJ. AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol.* 2018;19(2):121-135. doi:10.1038/nrm.2017.95
73. Luo H, Li Q, O'Neal J, Kreisel F, Le Beau MM, Tomasson MH. c-Myc rapidly induces acute myeloid leukemia in mice without evidence of lymphoma-associated antiapoptotic mutations. *Blood.* 2005;106(7):2452-2461. doi:10.1182/blood-2005-02-0734
74. Biver E, Thouverey C, Magne D, Caverzasio J. Crosstalk between tyrosine kinase receptors, GSK3 and BMP2 signaling during osteoblastic differentiation of human mesenchymal stem cells. *Mol Cell Endocrinol.* 2014;382(1):120-130. doi:10.1016/j.mce.2013.09.018
75. Banerji V, Frumm SM, Ross KN, et al. The intersection of genetic and chemical genomic screens identifies GSK-3 α as a target in human acute myeloid leukemia. *J Clin Invest.* 2012;122(3):935-947. doi:10.1172/JCI46465
76. Rubinfeld B, Albert I, Porfiri E, Fiol C, Munemitsu S, Polakis P. Binding of GSK3 β to the APC- β -catenin complex and regulation of complex assembly. *Science.* 1996;272(5264):1023-1026. doi:10.1126/science.272.5264.1023
77. Rayasam GV, Tulasi VK, Sodhi R, Davis JA, Ray A. Glycogen synthase kinase 3: more than a namesake. *Br J Pharmacol.* 2009;156(6):885-898. doi:10.1111/j.1476-5381.2008.00085.x
78. Soulet F, Bodineau C, Hooks KB, et al. ELA/APELA precursor cleaved by furin displays tumor suppressor function in renal cell carcinoma through mTORC1 activation. *JCI Insight.* 2020;5(14):e129070. doi:10.1172/jci.insight.129070
79. Shen Y, Liu S, Fan J, et al. Nuclear retention of the lncRNA SNHG1 by doxorubicin attenuates hnRNPC-p53 protein interactions. *EMBO Rep.* 2017;18(4):536-548. doi:10.15252/embr.201643139
80. Wald O, Shapira OM, Izhar U. CXCR4/CXCL12 axis in non small cell lung cancer (NSCLC) pathologic roles and therapeutic potential. *Theranostics.* 2013;3(1):26-33. doi:10.7150/thno.4922
81. Franco R, Pirozzi G, Scala S, et al. CXCL12-binding receptors expression in non-small cell lung cancer relates to tumoral microvascular density and CXCR4 positive circulating tumoral cells in lung draining venous blood. *Eur J Cardiothorac Surg.* 2012;41(2):368-375. doi:10.1016/j.ejcts.2011.05.009
82. Rodriguez-Lara V, Peña-Mirabal E, Baez-Saldaña R, et al. Estrogen receptor beta and CXCR4/CXCL12 expression: differences by sex and hormonal status in lung adenocarcinoma. *Arch Med Res.* 2014;45(2):158-169. doi:10.1016/j.arcmed.2014.01.001
83. Wang M, Lin T, Wang Y, et al. CXCL12 suppresses cisplatin-induced apoptosis through activation of JAK2/STAT3 signaling in human non-small-cell lung cancer cells. *Onco Targets Ther.* 2017;10:3215-3224. doi:10.2147/OTT.S133055
84. Pei S, Minhajuddin M, Adane B, et al. AMPK/FIS1-mediated mitophagy is required for self-renewal of human AML stem cells. *Cell Stem Cell.* 2018;23(1):86-100.e6. doi:10.1016/j.stem.2018.05.021
85. Bosc C, Broin N, Fanjul M, et al. Autophagy regulates fatty acid availability for oxidative phosphorylation through mitochondria-endoplasmic reticulum contact sites. *Nat Commun.* 2020;11(1):4056. doi:10.1038/s41467-020-17882-2
86. Levy JMM, Towers CG, Thorburn A. Targeting autophagy in cancer. *Nat Rev Cancer.* 2017;17(9):528-542. doi:10.1038/nrc.2017.53
87. Amaravadi R, Kimmelman AC, White E. Recent insights into the function of autophagy in cancer. *Genes Dev.* 2016;30:1913-1930. doi:10.1101/gad.287524.116
88. White E. Deconvoluting the context-dependent role for autophagy in cancer. *Nat Rev Cancer.* 2012;12:401-410. doi:10.1038/nrc3262
89. Rabanal-Ruiz Y, Otten EG, Korolchuk VI. mTORC1 as the main gateway to autophagy. *Essays Biochem.* 2017;61(6):565-584. doi:10.1042/EBC20170027
90. Altman JK, Sassano A, Kaur S, et al. Dual mTORC2/mTORC1 targeting results in potent suppressive effects on acute myeloid leukemia (AML) progenitors. *Clin Cancer Res.* 2011;17(13):4378-4388. doi:10.1158/1078-0432.CCR-10-2285

91. Mancinelli R, Carpino G, Petrunaro S, et al. Multifaceted roles of GSK-3 in cancer and autophagy-related diseases. *Oxid Med Cell Longev*. 2017;2017:4629495. doi:10.1155/2017/4629495
92. Mendel DBLA, Xin X, Louie SG, et al. In vivo anti-tumor activity of SU11248, a novel tyrosine kinase inhibitor targeting vascular endothelial growth factor and platelet-derived growth factor receptors: determination of a pharmacokinetic/pharmacodynamic relationship. *Clin Cancer Res*. 2003;9(1):327-337.
93. Wu M, Li C, Zhu X. FLT3 inhibitors in acute myeloid leukemia. *J Hematol Oncol*. 2018;11(1):133. doi:10.1186/s13045-018-0675-4